

MICROFLORA IN PREGNANCY

Pages with reference to book, From 79 To 81

Salar Zai (PMRC Research Centre, Khyber Medical College, Peshawar.)

Saeeda Majeed (Department of Obstetric and Gynaecology, Lady Reading Hospital, Peshawar.)

Johar Khatoon (University Town, Peshawar.)

Abstract

Three hundred high vaginal swabs from pregnant women were cultured to investigate the vaginal flora during pregnancy. Two hundred swabs obtained from a private clinic were cultured aerobically for bacteria only, the remaining 100 from antenatal clinic of Khyber Hospital Peshawar were cultured both aerobically and anaerobically for bacteria and fungi (candida). Vaginal smears from patients in antenatal clinic were also examined for the presence of *Trichomonas vaginalis*. The organisms isolated were *Streptococcus faecalis*, *Escherichia coli*, *Staphylococcus aureus*, *Streptococcus agalactiae*, *Pseudomonas aeruginosa* and *Candida albicans*. *Candida albicans* was the most common organism isolated. *Staphylococcus epidermidis* was the only pathogenic growth obtained. *Trichomonas vaginalis* was observed in only 5 smears. The only anaerobic growth was that of gram positive cocci. (JPMA 63 ; 79 1996).

INTRODUCTION

The vaginal flora vary considerably with the PH of the secretions and the amount of glycogen present in the epithelium. These factors in turn depend on ovarian function. The cervical canal is generally sterile or contains only a few bacteria, probably due to its relatively alkaline secretions.

The organisms present are identical to those found in the upper vagina. Bacteriological study of the female genital tract has shown that enterobacteriaceae and anaerobic bacteria are frequent pathogens. However the flora are variable. This study was undertaken to estimate the vaginal flora during pregnancy.

MATERIAL AND METHODS

The material consisted of high vaginal swabs from patients selected from the Antenatal clinic giving positive history of vaginal discharge and pruritus. Patients were put in the lithotomy position and vulva, cleaned with plain swab, Cusco's bivalve speculum was inserted in the vagina and swabs taken from the posterior fornix and immediately inserted in the glass tube. The swabs were cultured on to blood agar, MacConkey's agar, S.F. medium and Todd Hewitt broth. For the growth of *Candida albicans* the swabs were cultured on Sabouraud's dextrose agar medium.

Mother specimen of the vaginal discharge was taken with a sterilized loop and smear was made on a glass slide along with a drop of saline and covered with a cover slip. It was examined microscopically for the presence of trichomonal infection. Inoculated plates were incubated both aerobically and anaerobically for 18 hours at 37°C and the organisms isolated were identified by biochemical tests. The sensitivity test of the pathogenic organisms was performed by a paper disc diffusion method. The antibiotic discs included Nalidixic Acid, Gentamicin, Erythromycin, Penicillin and Carbencillin.

RESULTS

Three hundred high vaginal swabs were cultured. Two hundred from pregnant women attending a

private clinic, while the remaining from antenatal clinic of Khyber Hospital Peshawar. The ages of these varied from 17 to 38 years and the duration of pregnancy from 3 to 9 months. Vaginal smears from one hundred patients of antenatal clinic were also examined microscopically for trichomonal infection.

Three types of discharge was seen in these patients. Watery, thick white and thick creamy offensive discharge. The main complaints in these patients were embarrassing itching, malodorous discharge, burning, irritation and dysparunia.

The most common infective organisms isolated from the patients in antenatal clinic were *Escherichia coli* (18%), *Staphylococcus aureus* (26%), *Streptococcus agalactiae* (16%), *Streptococcus faecalis* (16%), *Pseudomonas aeruginosa* (46%) and *Staphylococcus epidermidis* (20%). Anaerobic streptococci were isolated from 5 cases, *Candida albicans* in 50 and *Trichomonas vaginalis* in 5 cases. *Staphylococcus aureus* was confirmed by coagulase test and *Streptococcus agalactiae* and *Streptococcus faecalis* by grouping sera. The drug sensitivity pattern is shown in the accompanying table. Gentamicin was the most effective drug for *Escherichia coli* followed by Nalidixic acid and carbencillin. *Pseudomonas aeruginosa* was most sensitive to Gentamicin and *Streptococcus agalactiae* (GPB) to penicillin. Other drugs in order of effectiveness were erythromycin and ampicillin (Table).

Table
% of Strains of Species of Bacteria Sensitive to Antimicrobials.

Organisms their number	Nalidixic Acid	Gentamicin	Erythromycin	Clindamycin	Penicillin	Carbencillin	Ampicillin
<i>Streptococcus faecalis</i> (50)	N.T.	15(30)	37(74)	28(56)	30(60)	N.T.	36(72)
<i>Escherichia coli</i> (54)	27(50)	42(77.7)	2(3.5)	1(1.8)	2(3.7)	15(27.7)	8(14.8)
<i>Staphylococcus aureus</i> (78)	4(5.1)	59(74.3)	45(57.6)	60(76.9)	7(8.9)	37(47.4)	16(20.5)
<i>Streptococcus agalactiae</i> (50)	N.T.	21(42)	41(82)	N.T.	46(92)	N.T.	33(66)
<i>Pseudomonas aeruginosa</i> (14)	12(85.7)	13(92.1)	1(7.1)	1(7.1)	1(7.1)	9(64.2)	1(7.1)

DISCUSSION

The vaginal microbial flora were often investigated during sixties.^{1,2} In seventies advances in microbiological techniques led to the isolation and identification of new bacterial species³. Detailed quantitative microbiological methods. also become feasible in the laboratory. Those new techniques have spurred to a series of investigations.⁴⁻¹¹

Afzal et al¹² reported a 33% incidence rate for *Staphylococcus aureus*, *Staphylococcus epidermidis* 26 percent, *Streptococcus pyogenes* 25%, *Escherichia coli* 22 percent and *Streptococcus faecalis* 18 percent. In the present survey *Streptococcus agalactiae* (GPB) was isolated however no *Streptococcus agalactiae* (GPA) streptococcal strain was found although specific culture medium (Todd Hewitt broth) for its cultivation and grouping sera for confirmation were used.

Marked clinical improvement was achieved with nystatin vaginal cream locally and Mycostatin oral tablets. High cure rate after a single course of therapy was obtained except in two cases, where relapses occurred and a second course of antibiotics was given. Not a single patient showed resistance to treatment or reinfection. It is suggested that consorts of these cases should also be checked as they may

be harbouring similar infective organisms.

REFERENCES

1. Morison, C.R. Antibiotic vaginitis 1966; Br. Med. J., 1: 291.
2. Morris, C.A. and Morris, D.F. Normal Vaginal microbiology of women of childbearing age in relation to the use of oral contraceptives and vaginal tampons. J. Clin. Pathol., 1967; 20 : 636-640.
3. Thadepalli, H., Gorbach, S.L. and Reith, L. Anaerobic infections of the female genital tract: bacteriologic and therapeutic aspects. J. Obstet. Gynaecol., 1973; 117: 1034.
4. Bartlett, J.G., Onderdonk, A.B., Drude, E., Goldstein, C., Anderka, M., Alpert, S. and McCormack, W.M. Quantitative bacteriology of the vaginal flora. J. Infect. Dis., 1977; 136:271.
5. Corbach, S.L., Menda, K.B., Thadepalli, H. and Keith, L. Anaerobic microflora of the cervix in healthy women. Am. J. Obstet. Gynaecol., 1973; 117:1053.
7. Levison, M.E., Corman, L.C., Carrington, E.R. and Kaye, D. Quantitative microflora of the vagina. Am. J. Obstet. Gynaecol., 1977; 127:80.
8. Lindner, J.G.E.M., Plantema, F.E.F. and Hoogkamp-Korstanje, J.A. Quantitative studies of the vaginal flora of healthy women and of obstetric and gynaecological patients. J. Med. Microbiol., 1978; 11:233.
9. Onderdonk, A.B., Polk, B.F., Moon, N.E., Goren, B. and Bartlett, J.G. Methods for quantitative vaginal flora studies. Am. J. Obstet. Gynaecol., 1977; 128:777.
10. Osborne, G.N., Wright, R.C. and Grubin, L. Genital bacteriology, comparative study of premenopausal women with postmenopausal women. J. Obstet. Gynaecol., 1979;135: 195.
11. Tashjian, J.H., Coulam, C.B. and Washington, J.A. Vaginal flora in asymptomatic women. Mayo Clin. Proc., 1976;51:557.
12. Afzal, H., Ijaz, M.K. and Akhtar, F. Bacteriological study of leukorrhoea. JPMA., 1982;32:163.